e-ISSN 2371-770X

Modern Medical Laboratory Journal

DOI: 10.30699/mmlj17.2.1.15

A Review of Mycoplasma in Laboratory Mice

Zahra Masoumalinejad ¹, Mohammad Reza Zinatizadeh*², Najmeh Tahmasebiabdar ¹

- 1. Dept. of Microbiology, Sirjan Branch, Islamic Azad University, Sirjan, Iran
- 2. Dept. of Genetics, Faculty of Science, Tonekabon Branch, Islamic Azad University, Mazandaran, Iran

Mycoplasma, Arthritidis, Muris, Neurolyticum, Pulmonis Article Info

Received 2019/05/21; Accepted 2019/06/26; Published Online 2019

ABSTRACT

Mycoplasma are small, cell-free bacteria enclosed by a membrane. These bacteria belong to the class of Mollicutes, the order of Mycoplasma tales, and the genus of Mycoplasma. There are more than 100 identified species of mycoplasma. The ratio of cytosine to guanine in its DNA is 23-40% and its genome size is 1350-600 kb. Mycoplasma require cholesterol to grow, and the temperature suitable for the growth of this bacteria is 37°C. Mycoplasma cause contamination and infections in humans and animals. Some mycoplasma species are seen only in animals. In general, mycoplasma are colonized at the surface of the mucus, and most species are noninvasive. Five main species of mycoplasma have been identified in laboratory mice, including: M.arthritidis, M.collis, M.muris, M.neurolyticum, and M.pulmonis. These species generally require protein-rich environments that contain 10-15% of the animal's serum, and their growth requires nicotinamide adenine nucleotide (NAD), which is commonly used to cultivate mycoplasma in mice. Laboratory research has found that mycoplasmas contamination has an adverse effect on animals. Therefore, it is important that health monitoring programs are implemented as a quality control for animals used in laboratory research.

Corresponding Information: Mohammad Reza Zinatizadeh, Dept. of Genetics, Faculty of Science, Tonekabon Branch, Islamic Azad University, Mazandaran, Iran, E-mail: zinati3333@gmail.com

Copyright © 2019. This is an open-access article distributed under the terms of the Creative Commons Attribution-noncommercial 4.0 International License which permits copy and redistribute the material just in noncommercial usages, provided the original work is properly cited.

Introduction

In 1898, Nocard and Roux separated certain bacteria from cattle with pneumonia (1). In 1929, Novak proposed the name of mycoplasma for these bacteria without cell walls, despite the split strands in production and reproduction. The name derives from, two words, the Greek word myco, meaning fungus, and the French word "plasma", meaning formed (2). By analyzing the sequence of the 16S rRNA gene, mycoplasma are thought to have been derived from gram-positive bacteria and clostridia around 600 million years ago, with a loss of the unnecessary parts of its genome (3).

Mycoplasma are small, Gram-negative, lack cell walls, and are enclosed by a membrane. These microorganisms grow relatively slowly and generally prefer the environment to be about 37-38° C. They are almost resistant to thallous acetate and penicillin, which are often used in culture

environments to postpone the growth of bacterial and fungal infections (4).

Five main species of mycoplasma have been identified in laboratory mice namely, *M. arthritidis*, *M. collis*, *M. muris*, *M. neurolyticum*, and *M. pulmonis*. Among these mycoplasma, *M. pulmonis* is responsible for one of the most common mycoplasma contaminations in mice and rats (5).

Sanchez et al. reported *M. pulmonis* to be an etiologic agent. A high count of these bacteria is often found in the ovaries, uterus, and respiratory systems in mice and rats (6).

Classification

Mycoplasma, as a member of the class Mollicutes, the order Mycoplasma tales, the genus of Mycoplasma, include more than 100 identified species (Fig 1). The ratio of cytosine to guanine in its DNA is 23-40% and its genome size is 1350–600 kb. It requires cholesterol to grow, and the

temperature suitable for the growth of these bacteria is 37°C (7).

The class Mollicutes covers more than 100 species of mycoplasma of plants and vertebrates, as well as insects. The order Mycoplasmatales is divided into three families of Mycoplasmataceae,

Acholeplasmatacea, and Spiroplasmatacea. The Mycoplasmataceae family consists of two genus Mycoplasma, of which more than 70 species have been identified, and many of which are pathogenic to humans and animals, and Ureaplasma, differentiated by urea hydrolysis (3,7,8).

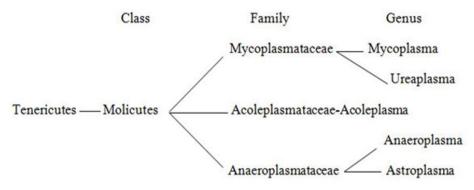


Fig1. Mycoplasma classification

Features of Mycoplasma in Laboratory Mice

In general, the mycoplasma species in mice require protein-rich environments that contain 10-15% of the serum. Other elements are derivatives of yeasts. The fermented carbohydrates include: 1. Those that ferment glucose 2. Those that do not ferment glucose (9).

Glucose is often added to the fluid medium for the growth of species that ferment glucose. Glucose is also a sign of growth. When glucose is fermented, it produces acid. So, in order to identify fermentation, phenol is added to the environment (8,3).

Phosphatase activity is often used in mycoplasma species that do not ferment glucose from arginine as an energy source, as shown in **Table 1**.

Table 1. Fermentation of Glucose an	l Arginine Hydro	olysis in Mycopi	lasma in Mice
-------------------------------------	------------------	------------------	---------------

Species	Usual host	Fermentation of Glucose	Arginine Hydrolysis
M.arthritidis	Mice	-	+
M.collis	Mice/Dog	+	-
M.muris	Mice	-	+
M.neurolyticum	Mice	+	-
M.pulmonis	Mice	+	-

Mycoplasma Contamination in Mice *Mycoplasma pulmonis*

This contamination is very common in laboratory mice and affects the middle ear space. Infection in the middle ear causes middle ear otitis media, which causes the neck to twist and deviate (10). This mycoplasma contamination causes severe respiratory problems in mice and laboratory rats, and it causes infection of the reproductive system in

mice (11). The prevalence of this disease among the experimental animals is between 20-60%. It also colonizes the trachea and throat, causing pneumonia and genital diseases, thus reducing reproductive rates (12). The transmission of the mycoplasma during fetal development occurs in two ways: by transmission through amniotic fluid or invasion of a pair and by transmission due to intrauterine infection or during implantation (13). In 1984, Furr

and Robinson found that, *the M. pulmonis* infection in the vaginal tract of TO and CBA mice was greatly cured with the help of hormonal therapy (progesterone), which was used to treat 33% of TO and 50% of RBC mice (14). In 2002, Barto et al. isolated bacteria from M. pulmonis from Mus musculus mice with symptoms of respiratory disease (15) In 2014, Shafaati et al., isolated the molecular identification of *M. pulmonis* from rat respiratory tract using PCR on the 16s rRNA gene, this being the first study in Iran to isolate *M. pulmonis* from the respiratory tract of rats (16).

Mycoplasma neurolyticum

In 1938, the bacteria were first isolated from the brain of the mice (17). In 1965, *M. neurolyticum* was isolated from the nasal mucous membranes and lungs of carrier animals, which showed no clinical symptoms (18). In 1979, Hill, studied *M. neurolyticum* in mice and rats, and the results showed that 78% of rats and 58% of mice with *M. neurolyticum* were infected (19). Taley, in 1981, described the bacteria as a mammalian brain organism that remains stressed and causes nerve disorders (20).

Mycoplasma collis

In 1983, these bacteria were isolated from the nasal cavity and conjunctiva of mice and rats for the first time (21). This species of mycoplasma grows in an environment of pH=7.8 at an optimal temperature of 35°C in anaerobic conditions. Some researchers describe this species of mycoplasma as mycoplasma in dogs, but *M. collis* was originally identified in rodents (22). So far, no accurate and complete report has been made on this mycoplasma species.

Mycoplasma muris

In 1983, McGarrity et al. a study based on the immune response mice led to the identification of the mycoplasma (23). In this study, all mice were pregnant and had tumors. The age group of the mice was three to 10 months old, which based on the morphological similarity with the mycoplasma, a new species of mycoplasma called Muris came into existence (23).

M. muris are small pathogenic bacteria that lives in the genital tract of female mice (23). Infection by M. muris may have harmful effects on the reproductive health of female mice (23). Weisburg

et al., based on the 16s rRNA gene, identified M. muris as the ancestors of the group of pneumonia, which consists of three distinct clusters of M. pneumonia, M. muris, and Ureaplasma urealyticum (24). Van Kuppeveld et al. designed specific primers for nine species of mycoplasma for humans and rodents (including the five species mentioned in this article) from 16s rRNA and evaluated them with the PCR test (25). In 2017, Zinatizadeh et al. identified this rare mycoplasma in NIH mice from Razi Vaccine and Serum Research Institute, Alborz, Iran. A total of 18% of the NIH rats were infected with M. muris in the Department of Animal Breeding, Razi Vaccine and Research Institute, and by using a phylogenetic analysis, a new species of M. muris was recorded in the gene bank (26).

Mycoplasma arthritidis

This mycoplasma infection is not common and is usually found in large laboratory mice. M. arthritidis causes arthritis of the joints in the mice Some researchers believe that (27).the microorganism enters the body through the mouth and mucous membranes, and there may be a latent infection (27). The clinical signs include swelling of the fingers and legs (28). This species of mycoplasma grows in a neutral pH medium (7.0) at an ideal temperature of 37°C, and is able to grow in the presence or absence of oxygen (29). The growth of *M. arthritidis* depends on the culture medium. In fiber tissue, it expands as a dense mass in the center, which requires sugar, proteins, amino acids, vitamins, and nucleic acids for growth (29).

Conclusion

Mycoplasma contamination has an adverse effect on laboratory animals, which interferes with the results of the researches conducted in laboratories. The presence of mycoplasma should be monitored for this reason. It is significant that many of the contaminated laboratory animals show no clinical symptoms. Therefore, it is important that health monitoring programs are implemented as a quality control for animals used in laboratory research.

Conflict of Interest

Authors declared no conflict of interest.

References

- Nocard E. Roux ER. Le microbe de la peripneumonie. Ann Inst Pasteur (Paris). 1898;12: 240-62.
- 2. NOWAK J. Morphologie, nature et cycle evolutif du microbe de la péripneumonie des bovidés. Ann inst Pasteur. 1929;43:1330-52.
- 3. Razin S. Adherence of pathogenic mycoplasmas to host cells. Biosci Rep. 1999;19(5):367-72.
- George M. Garrity Sc. D, Bergey's Manual Trust. Bergey's Manual of Systematic Bacteriology. 2nd Ed. USA: Department of Microbiology and Molecular Genetics Michigan State University East Lansing; 2005.
- Van Kuppeveld F, Van der Logt J, Angulo A, Van Zoest M, Quint W, Niesters H, et al. Genus-and species-specific identification of mycoplasmas by 16S rRNA amplification. Appl Environ Microbiol. 1992;58(8):2606-15.
- Sanchez S, Tyler K, Rozengurt N, Lida J. Comparison of a PCR-based diagnostic assay for Mycoplasma pulmonis with traditional detection techniques. Lab Anim. 1994;28(3):249-56.
- Razin S, Herrmann R, editors. Molecular biology and pathogenicity of mycoplasmas. New York: Kluwer Academic; 2002.
- 8. Razin S, Yogev D, Naot Y. Molecular biology and pathogenicity of mycoplasmas. Microbiol Mol Biol Rev. 1998;62(4):1094-156.
- De la Fe C, Assuncao P, Rosales R, Antunes T, Poveda J. Characterisation of protein and antigen variability among Mycoplasma mycoides subsp. mycoides (LC) and Mycoplasma agalactiae field strains by SDS-PAGE and immunoblotting. Vet J. 2006;171(3):532-8.
- Goto K, Yamamoto M, Asahara M, Tamura T, Matsumura M, Hayashimoto N, et al. Rapid identification of Mycoplasma pulmonis isolated from laboratory mice and rats using matrix-assisted laser desorption ionization time-of-flight mass spectrometry. J Vet Med Sci. 2012;74(8):1083-6.
- 11. Booth JL, Umstead TM, Hu S, Dybvig KF, Cooper TK, Wilson RP, et al. Housing conditions modulate the severity of Mycoplasma pulmonis infection in mice deficient in class A scavenger receptor. Comp Med. 2014;64(6):424-39.

- 12. Cassell GH, Clyde WA, Davis JK. Mycoplasmal respiratory infections. The mycoplasmas. 1985:65-106.
- CassellZ GH, Davis JK, Simeckaz JW, Lindsey JR, Cox NR, Rossz S, et al. Mycoplasmal infections: disease pathogenesis, implications for biomedical research, and control. Viral and Mycoplasmal of Laboratory Rodents: Effects on Biomedical Research. 2012;87.
- Furr PM, Taylor-Robinson D. Enhancement of experimental Mycoplasma pulmonis infection of the mouse genital tract by progesterone treatment. J Hyg (Lond). 1984;92(2):139-44.
- **15.** Barreto ML, Nascimento ERd, Campos CAdM, Nascimento MdGFd, Lignon GB, Lira MLF, et al. Detection of Mycoplasma pulmonis in laboratory rats. Braz J Microbiol. 2002;33(3):260-4.
- Shafaati M R, Khadem N, Yazdan setad S, Momeni T, zardadi M. Isolation and molecular identification of Mycoplasma pulmonis from the respiratory tract of Rattus rattus. Iran J Med Microbiol. 2016;9(4):24-31.
- 17. Findlay G, Maccallum F, Mackenzie R. Rolling disease: new syndrome in mice associated with a pleuropneumonia-like organism. The Lancet. 1938;232(6018):1511-3.
- 18. Adler H. Mycoplasmosis in animals. Adv Vet Sci. 1965;10:205-44.
- 19. Hill A. Mycoplasma isolation from the central nervous system. Vet Rec. 1979.
- 20. Tully J. Mycoplasmal toxins. Israel Isr J Med Sci. 1981;17(7):604-7.
- 21. Hill AC. Mycoplasma collis, a new species isolated from rats and mice. Int J Syst Bacteriol. 1983.33(4):847-51.
- 22. Johansson K-E, Pettersson B. Taxonomy of Mollicutes. In: Razin S, Herrmann R, editors. Molecular Biology and Pathogenicity of Mycoplasmas. New York: Kluwer; 2002.pp. 1–27.
- 23. McGarrity G, Rose D, Kwiatkowski V, Dion A, Phillips D, Tully J. Mycoplasma muris, a new species from laboratory mice. Int J Syst Evol Microbiol. 1983;33(2):350-5.
- 24. Weisburg W, Tully J, Rose D, Petzel J, Oyaizu H, Yang D, et al. A phylogenetic analysis of the mycoplasmas: basis for their classification. J Bacteriol. 1989;171(12):6455-67.

- 25. Van Kuppeveld F, Van Der Logt J, Angulo A, Van Zoest M, Quint W, Niesters H, et al. Genus-and species-specific identification of mycoplasmas by 16S rRNA amplification. Appl Environ Microbiol. 1993;59(2):655.
- Zinatizadeh M R, Abedini F, Jafarpour M, Masoumalinejad Z. Identification of Mycoplasma Muris Isolated from Vaginal Samples of NIH Mice . Mod Med Lab J. 2017;1(3):100-6.
- 27. Constantopoulos GE, McGarrity GJ. Activities of oxidative enzymes in mycoplasmas. J Bacteriol. 1987;169(5):2012-6.
- 28. Barden JA, Tully JG. Experimental arthritis in mice with Mycoplasma pulmonis. J Bacteriol. 1969;100(1):5-10.
- Gel'man NS, Lukoyanova MA, Ostrovskii DN. The respiratory chain of bacteria. Respiration and Phosphorylation of Bacteria: Springer; 1967. p. 71-159.

How to Cite This Article:

Masoumalinejad Z, Zinatizadeh M R, Tahmasebiabdar N. A Review of Mycoplasma in Laboratory Mice. Mod Med Lab J. 2019; 2 (2):127-131